Multiplex PCR Protocol

Modified from protocol by Giacomo Bernardi

- 1. Dilute stock primers from 1 mM to 50 uM.
 - a. Make and label tubes of 19 uL water.
 - b. Add 1 uL stock primer to each tube.
- 2. Make primer mix:
 - a. Each reaction requires 1uL of primer mix containing 0.04 uL of each primer.
 - b. For N reactions, mix (N*1.1)uL of primer and add water to make (N*1.1)uL total volume.
- 3. Add primer mix, master mix, and water to 96 well plate.
- 4. Add DNA to plate
- 5. Amplify in thermocycler using desired PCR parameters. For Amphiprion, I have been successful using: 95°/15'; 40x(94°C/30";57°C/90";72°C/60"); 72°C/7')
- 6. Mix 26.15uL size standard (ROX 500) with 973.85uL HiDi Formamide to make 1000uL of solution. Add 10uL of this solution to each well of a 96-well plate.
- 7. Add 0.5uL of PCR product to each well.
- 8. Seal and send out for analysis or do it yourself!

Notes:

Allow at least 2.5 hours for PCR to complete.